

Understanding the effects of grazing on cyanoHAB toxicity

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Freshwater ecosystems have been critical to sustaining life and establishing civilizations throughout history¹. **As global populations grow, increased urbanization, agricultural and industrial production, combined with poor sanitation practices, have led to a widespread increase in**



Figure 1: Remote Sensing Image of Lake Erie Bloom during Toledo Water Crisis in 2014.

pollution of freshwater bodies, creating an impending crisis of clean water scarcity in many regions of the world. One major threat to many freshwater systems is the proliferation of toxic algal and cyanobacterial blooms. Western Lake Erie has experienced several large toxic cyanobacterial harmful algal blooms (cyanoHABs) in the last few years, leading to major environmental and human health risks². Prior to P load reductions in the 1970s, cyanoHABs in Lake Erie were mostly N₂-fixers (e.g. *Aphanizomenon* and *Dolichospermum*³, formerly *Anabaena*⁴. Now, summer cyanoHABs are primarily non-N₂-fixing *Microcystis*, which blooms in Maumee Bay⁵ and *Planktothrix*, which blooms in Sandusky Bay⁶. The toxic *Microcystis* cyanoHAB in Lake Erie in August 2014 (Figure 1) created a water crisis that forced the shutdown of the public water supply to over 400,000 people in Toledo, OH². Blooms in Lake Erie have been occurring sporadically for decades, and have now become a regular occurrence⁶, and climate change increases the likelihood for more expansive blooms, exposing larger populations to water-borne toxins⁷. Despite cyanoHAB toxicity being a major human and ecosystem health hazard, it is not fully understood what causes and influences the underlying toxicity⁸. Several studies have investigated the effects of cyanotoxins on grazers^{9,10}, **but no studies have investigated the link between grazing activity and toxicity. It is hypothesized that, since cyanotoxins may negatively impacts grazer communities, there should be a feedback loop between grazers and cyanoHAB toxin production.** This study will investigate the impact of grazing rates on toxin production by utilizing a dilution experiment. Dilution experiments are used to quantify grazing rates in natural phytoplankton communities. The experiment will be run using bioassays following the methodology of Landry and Hassett¹¹ to quantify grazing and pair that with a cyanotoxin screening (e.g. Microcystin, anatoxin, etc.) using the methods indicated by Boyer¹². The filtered lake water used for the dilution will be filtered using a 0.2 μm pore-size filter to remove algae, cyanobacteria, and grazers¹³. All bioassays will receive a 20 mg L^{-1} NaHCO_3 addition to avoid the potential for inorganic C limitation¹⁴. The bioassays will then be run at 5%, 10%, 15%, 25%, 50%, 75%, and 100% dilutions in triplicate. Cyanotoxin synthesis will be calculated as the biomass-normalized change in cyanotoxin concentration and will be plotted against the dilution factor (Figure 2). This analysis will reveal if there is a correlation between the production of cyanotoxins and grazing, which will help fill gaps in the understanding of toxin production in cyanoHABs. **This research will help clarify what leads to differences in toxicity response in cyanoHAB bloom ecology and inform future research questions relating to mechanisms and interactions influencing cyanoHAB toxin dynamics.**

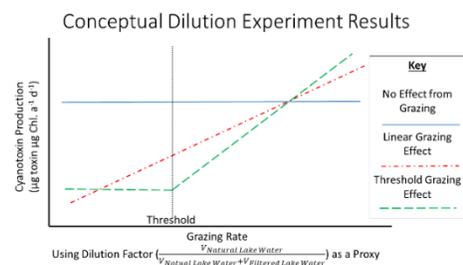


Figure 2: Conceptual Results from a Dilution Bioassay relating toxin response to grazing rate

Project Budget

Drive from Morehead City, NC to Put-In-Bay, OH (788 miles x ~\$0.50/mile)	\$400
Lodging	\$200
Sampling Supplies, Toxin Sample Shipping, and Toxin Analysis	<u>\$400</u>
Total Requested from Sigma Xi	\$1000

Total Project Budget (\$10,000)

Budget Justification

The drive from Morehead City, NC to Put-In-Bay, OH is 13.25 hours and lodging will be required for one night each way. Further transportation costs and lodging at the field site is covered by the Lake Erie Center grant. The sampling supplies refer to bioassay carafes that will be developed and made for this project. Most of the toxin analysis is covered by the Lake Erie Center grant mentioned; however, this project will require many more toxin screenings to occur, thus the \$400 will pay for the additional screenings and the shipping of the samples to the collaborator running the samples. The total project budget covers this component, boat time, lab technician time, drive back to Morehead City, NC, field site lab space, and field site lodging (please see Lake Erie Center Grant funding source).

Literature Citations

- (1) Haas M, Baumann F, Castella D, Haghypour N, Reusch A, Strasser M, Eglinton TI, Dubois N (2019) Roman-driven cultural eutrophication of Lake Murten, Switzerland. *Earth and Planetary Science Letters*.
- (2) Bullerjahn GS, McKay RM, Davis TW, Baker DB, Boyer GL, D'Anglada LV, Doucette GJ, Ho JC, Irwin EG, Kling CL, Kudela RM (2016) Global solutions to regional problems: collecting global expertise to address the problem of harmful cyanobacterial blooms. A Lake Erie case study. *Harmful Algae*.
- (3) Steffen MM, Belisle BS, Watson SB, Boyer GL, Wilhelm SW (2014) Review: Status, causes and consequences of cyanobacterial blooms in Lake Erie. *J. Great Lakes Research*.
- (4) Wacklin P, Hoffmann L, Komárek J (2009) Nomenclatural validation of the genetically revised cyanobacterial genus *Dolichospermum* (Ralfs ex Bornet et Flahault) comb. nova. *Fottea*.
- (5) Chaffin JD, Bridgeman TB & Bade DL (2013) Nitrogen constrains the growth of late summer cyanobacterial blooms in Lake Erie. *Advances in Microbiology*.
- (6) Davis TW, Bullerjahn, GS, Tuttle, T, McKay, RM, Watson, SB (2015) Effects of increasing nitrogen and phosphorus concentrations on phytoplankton community growth and toxicity during *Planktothrix* blooms in Sandusky Bay, Lake Erie. *Environ Sci Technol*.
- (7) Paerl HW & Huisman J (2009) Climate change: a catalyst for global expansion of harmful cyanobacterial blooms. *Environ. Microbiol. Rep.*
- (8) Smith ZJ, Martin RM, Wei B, Wilhelm SW, Boyer GL (2019) Spatial and Temporal Variation in Paralytic Shellfish Toxin Production by Benthic *Microseira (Lyngbya) wollei* in a Freshwater New York Lake. *Toxins*.
- (9) Engström, J, Viherluoto, M, & Viitasalo, M (2001). Effects of toxic and non-toxic cyanobacteria on grazing, zooplanktivory and survival of the mysid shrimp *Mysis mixta*. *Journal of Experimental Marine Biology and Ecology*
- (10) Van Donk, E, Cerbin, S, Wilken, S, Helmsing, NR, Ptacnik, R, & Verschoor, AM (2009). The effect of a mixotrophic chrysophyte on toxic and colony-forming cyanobacteria. *Freshwater biology*.
- (11) Landry MR and Hassett RP "Estimating the grazing impact of marine micro- zooplankton." *Marine biology*.
- (12) Boyer GL (2007) The occurrence of cyanobacterial toxins in New York lakes: Lessons for the MERHAB-lower Great Lakes program. *Lake Reservoir Management*.
- (13) Paerl HW and Davis TW, Pers. Comm.
- (14) Paerl HW, Hall NS, & Calandrino ES (2011). Controlling harmful cyanobacterial blooms in a world experiencing anthropogenic and climatic-induced change. *Science of the total environment*.